How effective is freezing at killing anisakid nematodes? An experimental evaluation of time-temperature conditions

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Conference of the West European **48th** Fish Technologists' Associatio Lisbon - Portugal, 15-18th October, 2018

The presence of parasites in marine fish is a serious problem for the fishing industry in many countries. Some species of parasite may pose a risk to consumers. The consumption of raw or inadequately cooked marine fish can lead to several disorders caused by the ingestion of viable anisakid nematodes. Anisakis simplex and A. pegreffii have been reported as causative agents of human infection, but other anisakids (Contracaecum osculatum and Pseudoterranova sp.) are also known to represent a potential hazard. Human

MATERIAL AND METHODS

Two species of fish were selected for the study:

• cod Gadus morhua - obtained from catches in the North Atlantic (Division 27.2.a.1 and 27.2.a.2);



health may also be compromised by allergic reactions to parasite antigens.

Treatment to kill viable parasites in fishery products for human consumption is mandatory in many countries. For parasites other than trematodes the freezing treatment must consist of lowering the temperature in all parts of the product to at least -20°C for not less than 24 h, or to -35°C for not less than 15 h (EU Regulation No. 1276/2011).

The question arises: how long should the product be held at the set freezing temperature (from the moment of placement to removal from the freezer) to meet the EU criteria?

The objective of the study

- to experimentally evaluate the time-temperature conditions necessary to kill anisakid larvae (A. simplex and Pseudoterranova sp.).

• herring *Clupea harengus* - caught in the southern Baltic Sea (Division 27.3.d, ICES Subdivision 24).

Two freezers with different rates of temperature change were used for the experiment:

- single-compressor freezer (model LGT-4725, Liebherr) with a conventional (static) cooling system (compressors with power of 433 W), without air circulation inside the refrigeration compartment;
- two-compressor freezer (model MDF-U443-PE, Panasonic) with a cascade cooling system (compressors with power of 400W and 750W) and fan-forced air circulation in the refrigerator for precise temperature uniformity.

FREEZING EXPERIMENT

(n = 240) were exposed to temperature of -15°C, -18°C, -20°C in the singlecompressor freezer (freezing rate 1.1°C/h) and -20°C, -25°C and -35°C in the twocompressor freezer (freezing rate 5.6°C/h).

During the entire freezing process, internal temperature of samples was recorded with wireless data loggers Track Sense Pro, Ellab.





RESULTS

Fig. 2. Fish samples in single-compressor (A) and two-compressor freezers (B).

The viability of anisakid larvae after freezing was assessed based on their mobility (EFSA 2010) and susceptibility to staining with malachite green (Leinemann and Karl 1988). Application of this dye allows dead and viable nematodes to be distinguished.



Fig. 3. Freezing curves for herring samples.

In total, the cod fillets contained 990 *Pseudoterranova* sp. larvae, while A. simplex larvae, which were also found in fillets, were less numerous (n = 72). All A. simplex and Pseudoterranova sp. larvae in cod fillets died at a temperature of -15°C or lower.

A total number of 1267 A. simplex larvae were found in herring body cavities, but not all were dead after freezing. Spontaneous movement was observed in 12



Fig. 4. Mobility and coloration of *A. simplex* larvae obtained from herring samples.

This research was supported by The National Centre for Research and Development under the Strategic Program Biostrateg (grant no. BIOSTRATEG2/296211/4/NCBR/2016).

parasites held in the single-compressor freezer for 24 h. Eight larvae kept at -15°C were active immediately after thawing. A few larvae survived at -18°C (two individuals) and -20°C (two individuals) and showed spontaneous mobility after thawing. All parasites stored in the single-compressor freezer at -20°C for 48 h and in the two-compressor device at $\leq -20^{\circ}C/24$ h were motionless after thawing. Stimulation with tweezers and incubation at 37°C did not provoke any activity.

Obtained results clearly demonstrate that the appropriate freezing process is essential to ensure the safety of fish products.

References: EFSA Panel on Biological Hazards (BIOHAZ) (2010) Scientific Opinion on risk assessment of parasites in fishery products. EFSA Journal 8(4): 1543 [91 pp]. Available online www.efsa.europa.eu.

Leinemann M, Karl H (1988) Untersuchungen zur Diferenzierung lebender und toter Nematodenlarven (Anisakis sp.) in Heringen und Heringenzeugnissen. Arch Lebensmittelhyg 39: 147-150.













