

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/323991635>

# Impact of hyperbaric oxygenation on oxidative stress in diabetic patients

Article in *Undersea & hyperbaric medicine: journal of the Undersea and Hyperbaric Medical Society, Inc* · January 2018

CITATIONS

0

READS

18

9 authors, including:



Anica Petkovic

University of Kragujevac

8 PUBLICATIONS 4 CITATIONS

SEE PROFILE



Ivan Srejavic

University of Kragujevac

54 PUBLICATIONS 68 CITATIONS

SEE PROFILE



Nevena Jeremic

University of Louisville

81 PUBLICATIONS 244 CITATIONS

SEE PROFILE



Jelena Bradic

COM DEV International

7 PUBLICATIONS 8 CITATIONS

SEE PROFILE

Some of the authors of this publication are also working on these related projects:



The effects of homocysteine and homocysteine- related compounds on cardiovascular system: the role of gasotransmitters NO, H2S and CO [View project](#)

Research Article

## Impact of hyperbaric oxygenation on oxidative stress in diabetic patients

Sandra Tepić, PhD <sup>1</sup>; Anica Petković, Assist. <sup>2</sup>; Ivan Srejšević, Assist. Prof. <sup>3</sup>;  
Nevena Jeremić, PhD Assist. Prof. <sup>2</sup>; Vladimir Živković, Assist. Prof. <sup>3</sup>;  
Slobodan Lončarević, Assist. Prof. <sup>4</sup>; Jovana Bradić, Assist. <sup>2</sup>; Vladimir Jakovljević, Prof. <sup>3,5</sup>;  
Miodrag Živković, MD <sup>6</sup>

- <sup>1</sup> Department of Urgent Medicine, Clinical Center “Zvezdara,” Belgrade, Serbia
- <sup>2</sup> Department of Pharmacy, Faculty of Medical Sciences, University of Kragujevac, Kragujevac, Serbia
- <sup>3</sup> Department of Physiology, Faculty of Medical Sciences, University of Kragujevac, Kragujevac, Serbia
- <sup>4</sup> Department of Dentistry, Faculty of Medical Sciences, University of Kragujevac, Kragujevac, Serbia
- <sup>5</sup> Department of Human Pathology, 1st Moscow State Medical University IM Sechenov, Moscow, Russia
- <sup>6</sup> HBO Medical Center, Belgrade, Serbia

CORRESPONDING AUTHOR: Prof. Vladimir Lj. Jakovljevic, MD, PhD – [drvladakbg@yahoo.com](mailto:drvladakbg@yahoo.com)

---

### ABSTRACT

Taking into consideration that high concentration of oxygen can express toxic effects due to production of reactive oxygen species (ROS), the aim of our investigation was to establish the influence of hyperbaric oxygenation on oxidative stress parameters and antioxidant enzymes in patients with diabetes mellitus (DM) type 2. Investigation included 50 patients with DM type 2 divided into two groups. The first group consisted of 25 patients, mean age 70 years, mean duration of illness 12 years and without manifest peripheral vascular complications (Wagner 0). The second group consisted of 25 patients, mean age 74 years, mean duration of illness 17 years and with manifest peripheral vascular complications (Wagner 1-5).

All patients underwent the same therapeutic protocol, which included 10 hyperbaric oxygenation therapies, once a day for a duration of 60 minutes, with an average partial oxygen pressure of 1.7 atmospheres absolute (ATA). In blood samples the following parameters of redox balance were determined: levels of nitrites (NO<sub>2</sub><sup>-</sup>), index of lipid peroxidation (TBARS), superoxide anion radical (O<sub>2</sub><sup>-</sup>), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and antioxidant enzymes superoxide dismutase (SOD) and catalase (CAT).

Our results clearly show that hyperbaric oxygen (HBO<sub>2</sub>) therapy does not have a pro-oxidative effect. Additionally, it seems that this procedure strongly mobilized the antioxidant enzyme system, thus improving defense from oxidative damage. All significant data are marked as P<0.05. Our results have shown that in terms of ROS production, HBO<sub>2</sub> can be safe to use in patients suffering from DM type 2 with or without vascular complications.

### INTRODUCTION

Diabetes mellitus (DM) is a progressive metabolic disorder characterized by hyperglycemia resulting from lack of insulin secretion or action. This in turn leads to disorders of lipid, protein and carbohydrate metabolism. According to absolute or relative deficiency of insulin, diabetes is classified as type 1 and type 2 [1]. DM is associated with cardiovascular complications, nephropathy, neuropathy and retinopathy due to glucose uptake regardless of the presence of insulin. In developed countries, this disease is an important source of mortality and morbidity [2]. The World Health Organization (WHO) has assessed that 346 million people have diabetes; this number is predicted to double by 2030 [3].

It has been proposed that one of the crucial events in the pathogenesis of diabetes mellitus is the extent of release of ROS (reactive oxygen species). There is also additional evidence that damage caused by free radicals is possibly involved in beta-cell destruction [4]. Persistent hyperglycemia is present in both types of diabetes and causes increased production of oxygen free radicals. Mechanisms responsible for production of free radicals imply autooxidation of glucose, glycosylation of protein and increased lipid peroxidation. Production of free radicals leads to oxidative stress, which is associated with several health complications [5]. Reduction of glutathione concentration leads to disruptions in cellular signaling and upregulation of NF-κB trans-

---

KEYWORDS: diabetes mellitus; hyperbaric oxygen treatment; redox status

cription, leading to oxidative conditions [6]. In cases of catalase deficiency,  $\beta$ -cells undergo oxidative stress by producing excess ROS, which leads to  $\beta$ -cells dysfunction and ultimately diabetes. The antioxidant enzymes also play an important protective role in the pathogenesis of diabetes mellitus [4].

Hyperbaric oxygen (HBO<sub>2</sub>) therapy is a therapeutic modality which provides extra oxygen to tissues. HBO<sub>2</sub> has been suggested as valuable addition to conventional treatment for many indications (e.g., decompression sickness, gas or air embolism, carbon monoxide poisoning, clostridial myositis and myonecrosis, arterial insufficiencies, osteomyelitis, diabetic foot ulcer) [7]. During HBO<sub>2</sub> therapy a person breathes pure oxygen (100%) at pressures greater than 1 atmosphere, usually 2 to 3 atmospheres absolute (ATA). By increasing blood oxygen content, HBO<sub>2</sub> creates a favorable gradient for the diffusion of oxygen into the tissues. In hypoxic tissues, the enhanced oxygen supply has multiple effects: a direct antihypoxic effect, a reduction in plasma viscosity, an increase in elasticity of erythrocytes, and a reduction of platelet aggregation. Furthermore, this treatment stimulates neocapillarization by increasing capillary penetrance of oxygen to about three times, and allows for nitric oxide (NO) production, thus supporting normal endothelium function [8]. HBO<sub>2</sub> is usually used in DM for the treatment of diabetic foot and peripheral vascular disease [9].

Considering the standpoint that high concentrations of oxygen can cause toxic effects due to production of ROS, the aim of our investigation was to establish the influence of hyperbaric oxygenation on oxidative stress parameters and antioxidant enzymes in patients with diabetes mellitus type 2 (DM2).

## MATERIALS AND METHODS

### Subjects

The investigation included 50 patients with DM2 who were divided into two groups classified according to the Wagner scale. The first group consisted of 25 patients, mean age 70 years, mean duration of illness 12 years and without manifest peripheral vascular complications (Wagner 0). The second group included 25 patients, mean age 74 years, mean duration of illness 17 years and with manifest peripheral vascular complications (Wagner 1-5). The study protocol was

approved by the Ethical Committee of the Faculty of Medical Sciences, University of Kragujevac, and it was conducted in accordance with the principles of the Declaration of Helsinki. All the participants were informed about the research protocol before giving their written consent to participate in the study.

### Hyperbaric oxygen therapy protocol

Hyperbaric treatment was performed at HBO Medical Center in Belgrade. All patients underwent the same therapeutic protocol, which included 10 hyperbaric oxygenation treatments, once daily with a duration of 60 minutes, with an average partial pressure of oxygen of 1.7 ATA. Therapy was conducted in monoplace hyperbaric chambers, BKL-S 303.

### Sample collection protocol

Blood samples for biochemical analysis were collected before treatment and then on the third, fifth, seventh and tenth day of treatment. After centrifugation of heparinized venous blood, plasma and erythrocytes were separated. Routine laboratory analyses were measured pre- and post-HBO<sub>2</sub>, including: blood glucose levels (BGL), glycosylated hemoglobin (HbA1c), urea, creatinine, creatine kinase and electrolytes such as potassium chloride and sodium chloride. These parameters of redox balance were then determined: levels of nitrites (NO<sub>2</sub>-), index of lipid peroxidation (TBARS), superoxide anion radical (O<sub>2</sub>-), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), and antioxidant enzymes superoxide dismutase (SOD) and catalase (CAT).

### Biochemical assays

Blood samples were drawn from an antecubital vein into a vacutainer test tube containing sodium citrate anticoagulant. Blood samples were analyzed immediately. Blood was centrifuged to separate plasma and red blood cells (RBCs). Biochemical parameters were measured spectrophotometrically, using a UV-1800 Shimadzu UV Spectrophotometer.

### Superoxide anion radical determination

The level of superoxide anion radical (O<sub>2</sub>-) was measured using nitro blue tetrazolium (NBT) reaction in TRIS-buffer combined with plasma samples and read at 530 nm [10].

**Hydrogen peroxide determination**

The protocol for measurement of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) is based on oxidation of phenol red in the presence of horseradish peroxidase [11]. A 200- $\mu$ l sample with 800  $\mu$ l of PRS (phenol red solution) and 10  $\mu$ l of POD (horseradish peroxidase) were combined (1:20). The level of H<sub>2</sub>O<sub>2</sub> was measured at 610 nm.

**Nitric oxide determination**

NO decomposes rapidly to form stable metabolite nitrite/nitrate products. Nitrite (NO<sub>2</sub><sup>-</sup>) was determined as an index of nitric oxide production with Griess reagent [12]. Then 0.1 mL 3 N PCA (perchloric acid), 0.4 mL 20 mM ethylenediaminetetraacetic acid (EDTA), and 0.2 mL plasma were put on ice for 15 minutes, then centrifuged 15 minutes at 6,000 rpm. After pouring off the supernatant, 220  $\mu$ l K<sub>2</sub>CO<sub>3</sub> was added. Nitrites were measured at 550 nm. Distilled water was used as a blank probe.

**Index of lipid peroxidation: thiobarbituric acid reactive substances**

The degree of lipid peroxidation in plasma was estimated by measuring thiobarbituric acid reactive substances (TBARS) using 1% thiobarbituric acid (TBA) in 0.05 NaOH, incubated with plasma at 100°C for 15 minutes and read at 530 nm. Distilled water was used as a blank probe. TBA extract was obtained by combining 0.8 mL of plasma and 0.4 mL trichloroacetic acid (TCA), then samples were put on ice for 10 min, and centrifuged for 15 minutes at 6,000 rpm. This method was described previously [13].

**Determination of antioxidant enzymes SOD and CAT**

Isolated RBCs were washed three times with three volumes of ice-cold 0.9 mmol/L NaCl, and hemolysates containing about 50 g Hb/L (prepared according to McCord and Fridovich [14] were used for the determination of CAT activity. CAT activity was determined according to Beutler [15]. Then 50  $\mu$ l of CAT buffer, 100  $\mu$ l hemolysate, and 1 mL 10 mM H<sub>2</sub>O<sub>2</sub> were added to the samples. Detection was performed at 360 nm. Distilled water was used as a blank probe. SOD activity was determined by the epinephrine method of Misra and Fridovich [16]. A 100- $\mu$ l hemolysate and 1-mL carbonate buffer were mixed, and then 100  $\mu$ l of epinephrine were added. Detection was performed at 470 nm.

**Statistical analysis**

IBM SPSS Statistics 20 was used for statistical analysis. Values were expressed as mean  $\pm$  SE. The Student's t-test and analysis of variance (ANOVA) were used for comparison between groups and inside groups. P-values lower than 0.05 were considered to be significant.

**RESULTS**

There were no statistically significant changes in BGL, Hb1Ac, urea, creatinine, creatine kinase and levels of electrolytes after HBO<sub>2</sub> compared to baseline values (Table 1).

**Levels of nitrites**

During the observed 10-day period of HBO<sub>2</sub> therapy, there were statistically significant changes in NO<sub>2</sub><sup>-</sup> between both groups. In the group with vascular complications a significant increase in this parameter was noticed on the third day at the beginning of therapy. Values of NO<sub>2</sub><sup>-</sup> decreased statistically significantly (P-values lower than 0.05 were considered to be significant) at the fifth day compared to the third day. Values of NO<sub>2</sub><sup>-</sup> in the group without vascular complications were increasing until the end of treatment. A statistically significant increase was seen in values of NO<sub>2</sub><sup>-</sup> measured on the 10th day versus its values measured on the seventh day (Figure 1A).

**Levels of index of lipid peroxidation measured as thiobarbituric acid reactive substances**

During the observed 10-day period of HBO<sub>2</sub> therapy, there were no statistically significant changes of TBARS values between both groups. However, values of TBARS in the group without vascular complications were lower compared to the other group. On the other hand, in the group with vascular complications, values of TBARS significantly differed between the fifth and seventh days, when values of this parameter increased (Figure 1B).

**Levels of superoxide anion radical**

Values of superoxide anion radical were statistically significantly increased in diabetic patients with vascular complications at 10th day compared to baseline values. However the highest values of this parameter were noticed on 3rd day of therapy. We have also found that levels of O<sub>2</sub><sup>-</sup> were decreased after HBOT compared to baseline values in group without vascular complications (Figure 1C).

**TABLE 1. The effects of HBO<sub>2</sub> on laboratory parameters in patients with DM type 2 with and without vascular complications**

| laboratory parameter                                    | Wagner 0                |                        | Wagner 1-5              |                        |
|---|-------------------------|------------------------|-------------------------|------------------------|
|   | before HBO <sub>2</sub> | after HBO <sub>2</sub> | before HBO <sub>2</sub> | after HBO <sub>2</sub> |
| <b>glucose</b> [mmol/L]<br>(ref. values: 4.1 – 5.9)     | 5.1                     | 5.0                    | 5.4                     | 4.9                    |
| <b>urea</b> [mmol/L]<br>(ref. values: 2.5 – 7.5)        | 10.2                    | 10.0                   | 10.3                    | 10.1                   |
| <b>creatinine</b> [μmol/L]<br>(ref. values: 62 – 115)   | 113                     | 109                    | 110                     | 108                    |
| <b>potassium</b> [mmol/L]<br>(ref. values: 3.5 – 5.1)   | 4.9                     | 4.8                    | 5.1                     | 5.0                    |
| <b>sodium</b> [mmol/L]<br>(ref. values: 136 – 148)      | 145                     | 144                    | 142                     | 144                    |
| <b>chlorides</b> [mmol/L]<br>(ref. values: 98 – 111)    | 101                     | 95                     | 98                      | 105                    |
| <b>creatine kinase</b> [U/L]<br>(ref. values: 32 – 300) | 416                     | 401                    | 420                     | 396                    |
| <b>HbA1c</b> [mmol/mol]<br>(ref. values: 24 – 43)       | 50                      | 45                     | 57                      | 49                     |

**Levels of hydrogen peroxide**

During the observed 10-day period of HBO<sub>2</sub> therapy, there were no statistically significant change in H<sub>2</sub>O<sub>2</sub> between groups. Baseline values in the group without vascular complications were significantly higher than values during 10 days of HBO<sub>2</sub>. We saw a significant decrease of this parameter on the third day and a significant increase on the seventh day of therapy in the group without vascular complications. In the group with vascular complications, values of this parameter showed a continuous decrease (Figure 1D).

**Superoxide dismutase activity**

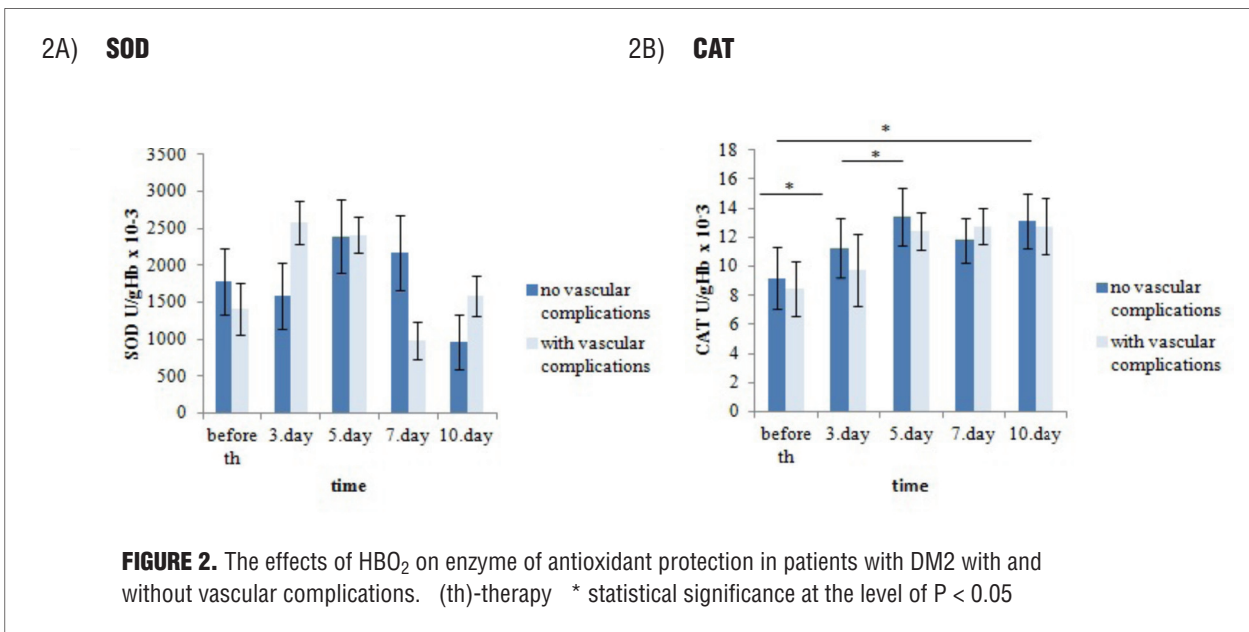
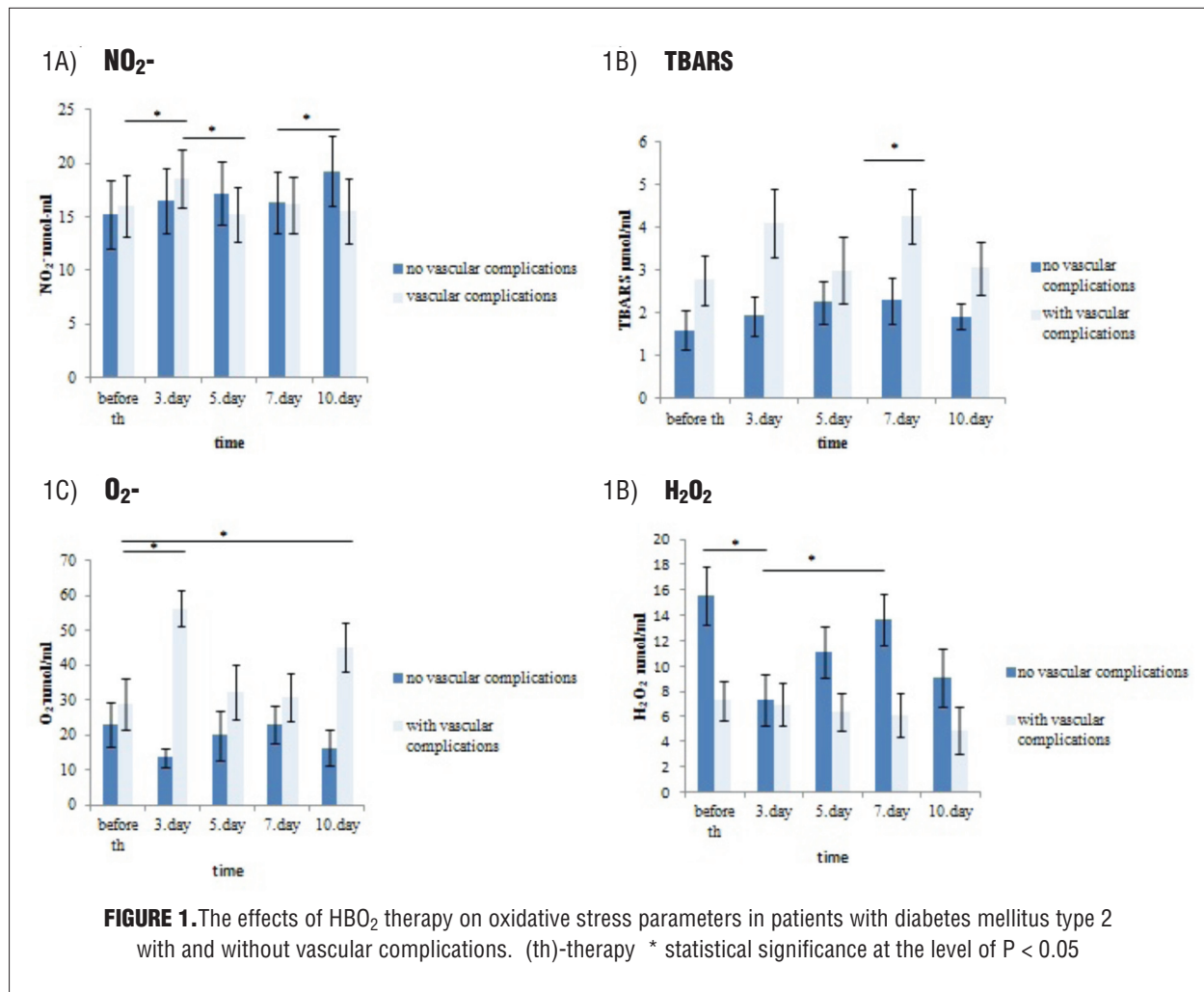
During the observed 10-day period of HBO<sub>2</sub> therapy, there were no statistically significant changes in SOD values between groups. In the group without vascular complications the values of SOD were high until the fifth day and then began decreasing until the end of HBO<sub>2</sub> treatment. However SOD activity in the group with vascular complications were lowest at the seventh day in comparison to values on the other days (Figure 2A).

**Catalase activity**

During the observed 10-day period of HBO<sub>2</sub> therapy, there were statistically significant changes of CAT values between groups. In the group without vascular complications, values of this parameter increased statistically significantly on the third, fifth and 10th day from the beginning of treatment. On the other hand, continuous increase of CAT was noticed in the group with vascular complications (Figure 2B).

**DISCUSSION**

Numerous studies have underlined the controversy that exists regarding the effect of HBO<sub>2</sub> on oxidative stress and enzymes of antioxidative defense in several pathophysiological models. Using healthy rats, a study by Ay and co-authors has shown a pro-oxidative activity of HBO<sub>2</sub> as a result of an increased index of lipid peroxidation (TBARS) and activity by SOD and CAT enzymes. In this study one HBO<sub>2</sub> treatment was applied for 90 minutes [17]. Otec and co-workers found that the pro-oxidative effect of HBO<sub>2</sub> is time-dependent [18]. Furthermore, HBO<sub>2</sub> reduced infarct size and



increased catalytic activity of CAT in an experimental infarct of the myocardium [19]. As regards clinical studies, some of them support HBO<sub>2</sub> treatment in different pathophysiological states while others not [20-24].

Chronic vascular complications are the main causes of mortality and morbidity in diabetic patients [25]. The main role in the development of chronic vascular complication is endothelial dysfunction, which leads to production of vasoconstrictors via oxidative degradation of NO and decreased synthesis of NO. Increased production of O<sub>2</sub><sup>-</sup> interacts with NO, producing one of the most harmful free radicals, peroxynitrite (ONOO<sup>-</sup>), which leads to vasoconstriction and hypoxia [26].

First, we examined the effects of HBO<sub>2</sub> on levels of pro-oxidants, such as NO<sub>2</sub><sup>-</sup> and TBARS in patients with DM2 without and with vascular complications. Our study has shown continuous increase of NO<sub>2</sub><sup>-</sup> values in diabetic patients without vascular complications. Also, values of this parameter were higher at the end of the therapy in comparison to the beginning of HBO<sub>2</sub> treatment.

On the other hand, in the group with vascular complications values of NO<sub>2</sub><sup>-</sup> were lower at the end of the HBO<sub>2</sub> in comparison to the beginning. Studies dealing with influence of HBO<sub>2</sub> on NO bioavailability show controversial results. Some of them have shown that HBO<sub>2</sub> acts via activation of nitric oxide synthase (NOS) [27]. Other studies have shown opposite effects of HBO<sub>2</sub> in the reduction of NO bioavailability [28].

An investigation of Chen and co-workers found that patients with DM2 had significantly increased levels of NO after HBO<sub>2</sub>. Our results as well as those of aforementioned studies could be consequence of high insulin levels, which might have a main role in NO production [29].

We found that the values of TBARS in the group without vascular complication slightly increased from the beginning of HBO<sub>2</sub> treatment until the seventh day. However, in a group with vascular complications the values of this parameter statistically increased on the third day. Literature data showed that indices of lipid peroxidation (TBARS) were higher in diabetic patients than in healthy patients who received HBO<sub>2</sub> therapy. As regards diabetic patients with vascular complications the levels of this parameter and allene oxide synthase (AOS) enzyme were much higher than

in patients without vascular complications [30-31]. Similar results were noticed in a study by Grudol and co-authors, where values of the index of lipid peroxidation in diabetic patients with terminal complications were high at the beginning of therapy and then unchanged 15 days after therapy [32].

Furthermore, other pro-oxidants, such as the superoxide anion radical and hydrogen peroxide were also significantly affected by HBO<sub>2</sub>. We have found decreased levels of O<sub>2</sub><sup>-</sup> after HBO<sub>2</sub> compared to baseline values in group without vascular complications, while levels of this parameter were higher in group with vascular complication. This indicates that patients with vascular complications have a lower activity of antioxidant enzymes in comparison to patients without vascular complications. Decreased levels of H<sub>2</sub>O<sub>2</sub> in the group with vascular complications might be due to increased CAT activity, which detoxifies hydrogen peroxide to water. Furthermore, one study revealed that levels of H<sub>2</sub>O<sub>2</sub> reduced due to increased activity by SOD and CAT, which are the major scavenging enzymes [33].

It is thought that HBO<sub>2</sub> therapy may increase oxidative stress via the production of reactive oxygen species, but literature data indicate that although oxidative stress can occur with HBO<sub>2</sub> treatment, it appears to be less of a concern at hyperbaric pressures under 2.0 ATA [34]. Our results clearly show that HBO<sub>2</sub> does not have a pro-oxidative effect. Parameters of oxidative stress were smoothly decreased during the treatment, which is in compliance with our previous study [35].

In order to complete a picture of the influence of HBO<sub>2</sub> on redox status we examined the activity of antioxidant enzyme systems such as SOD and CAT. Considering the fact that SOD is an enzyme that catalyzes the transformation of the superoxide anion radical to hydrogen peroxide, we can note that these two parameters are in correlation in both groups. Increase of O<sub>2</sub><sup>-</sup> is followed by decrease in SOD, particularly in the group with vascular complications. In the group without vascular complications, SOD values were high until the fifth day and then decreased until the end of HBO<sub>2</sub> treatment. These findings may suggest that HBO<sub>2</sub> should be used from the fifth day in diabetic patients without vascular complications, while therapy for patients who have diabetes with vascular complications should last longer.

As regards values of CAT during 10 days of HBO<sub>2</sub>

treatment, there was significant change in both investigated groups. Similar to SOD values, CAT levels in the group without vascular complications were high until the fifth day. High CAT values were followed by a drop in H<sub>2</sub>O<sub>2</sub> values particularly in the group with vascular complications. These results are in agreement with the results of previously conducted research that showed that hyperbaric oxygen treatment below 2.0 ATA can increase the activity of antioxidant enzymes including SOD and CAT [35].

There were no statistically significant changes in BGL, Hb1Ac, urea, creatinine, creatine kinase and levels of electrolytes after HBO<sub>2</sub>. This is not in accordance with the previously published papers in this field. Several studies suggest an increase in insulin production or modification of metabolism favorable to the diabetic, resulting in lower blood sugar. However, most of these studies have included diabetes mellitus type 1 (DM1) patients [36-38]. The possible mechanisms of hypoglycemia involve an increase in activity of insulin receptor sites and changes in insulin sensitivity due to upregulation of PPAR- $\gamma$  signaling associated with hyperbaric oxygen [39]. There are data that show insulin sensitivity increased within three days of hyperbaric oxygen treatment and maintained for 30 sessions [40].

Few studies have revealed that hypoglycemia might be encountered after HBO<sub>2</sub> among DM1 and DM2 patients [41-43]. The possible reason for a discrepancy between our results and results of mentioned authors may be duration of HBO<sub>2</sub> treatment. The shorter period of exposure to hyperbaric oxygenation present in our study might have not been sufficient to cause changes in parameters. There are limited data regarding effects of HBO<sub>2</sub> on kidney function and levels of electrolytes on the human population. Our

results are in agreement with literature data that HBO<sub>2</sub> does not cause renal impairment in a rat model [44,45], which reinforces the assumption that HBO<sub>2</sub> is safe in healthy rats.

Of course, better information about oxidative stress during HBO<sub>2</sub> treatment in such patients could be more scientifically significant if patients can be divided in several groups, especially those with vascular complications according to the Wagner scale.

A limitation of our study is a small sample size. Regarding that, we compared only effects in groups of patients with versus without general vascular complications.

## CONCLUSION

Our study has shown that in terms of the production of reactive oxygen species, hyperbaric oxygen therapy can be safe to use in patients suffering from diabetes mellitus type 2 with or without vascular complications. Moreover, it seems that this procedure strongly mobilizes the antioxidant enzyme system, thus improving defense from oxidative damage. Based on our results, treatment for patients without vascular complications should last five days; for diabetics with vascular complications treatment should last longer than five days. However, more research is required to establish the complete mechanism by which HBO<sub>2</sub> therapy can modify oxidation-reduction reactions in patients with DM2 in order to become an additional routine therapeutic strategy in the treatment of diabetes. Keeping in mind that HBO<sub>2</sub> was not connected to the changes in BGL, its usage can be considered safe in terms of this parameter. ■

*The authors declare that no conflicts of interest exist with this submission.*



## REFERENCES

1. Rayburn WF. Diagnosis and classification of diabetes mellitus: highlights from the American Diabetes Association. *J Reprod Med.* 1997; 42:585-586.
2. Shrivastava SR, Shrivastava PS, Ramasamy J. Role of self-care in management of diabetes mellitus. *J Diabetes Metab Disord.* 2013; 12:14.
3. World health organization: Diabetes – Factsheet. 2012. <http://www.who.int/mediacentre/factsheets/fs312/en/index.html>.
4. Szaleczky E, Prechl J, Fehér J, Somogyi A. Alterations in enzymatic antioxidant defence in diabetes mellitus--a rational approach. *Postgrad Med J.* 1999; 75:13-17.
5. Matough FA, Budin SB, Hamid ZA, Alwahaibi N, Mohamed J. The role of oxidative stress and antioxidants in diabetic complications. *Sultan Qaboos Univ Med J.* 2012; 12:5-18.
6. Pitocco D, Tesauro M, Alessandro R, Ghirlanda G, Cardillo C. Oxidative stress in diabetes: implications for vascular and other complications. *Int J Mol Sci.* 2013; 14:21525-21550. Review.
7. Stoekenbroek RM, Santema TB, Legemate DA, Ubbink DT, van den Brink A, Koelemay MJ. Hyperbaric oxygen for the treatment of diabetic foot ulcers: a systematic review. *Eur J Vasc Endovasc Surg.* 2014; 47:647-655.
8. Health Quality Ontario. Hyperbaric oxygen therapy for non-healing ulcers in diabetes mellitus: an evidence-based analysis. *Ont Health Technol Assess Ser.* 2005; 5:1-28.
9. Chen SJ, Yu CT, Cheng YL, Yu SY, Lo HC. Effects of hyperbaric oxygen therapy on circulating interleukin-8, nitric oxide, and insulin-like growth factors in patients with type 2 diabetes mellitus. *Clin Biochem.* 2007; 40:30-36.
10. Auclair C, Voisin E. Nitroblue tetrazolium reduction. In: Greenvald RA (ed) *Handbook of methods for oxygen radical research.* CRC Press, Boca Raton, 1985:123-132.
11. Pick E, Keisari Y. A simple colorimetric method for the measurement of hydrogen peroxide produced by cells in culture. *J Immunol Methods* 1980; 38:161-170
12. Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Analysis of nitrate, nitrite and [15N]nitrate in biological fluids. *Anal Biochem.* 1982; 126:131-138.
13. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem.* 1979; 95:351-358.
14. McCord JM, Fridovich I. The utility of superoxide dismutase in studying free radical reactions. I. Radicals generated by the interaction of sulfite, dimethyl sulfoxide, and oxygen. *J Biol Chem.* 1969; 244:6056-6063.
15. Beutler E. Catalase. In: Beutler E (ed) *Red cell metabolism, a manual of biochemical methods.* Grune and Stratton, New York, 1982:105-106.
16. Misra HP, Fridovich I. The role of superoxide-anion in the autooxidation of epinephrine and a simple assay for superoxide dismutase. *J Biol Chem.* 1972; 247:3170-3175.
17. Ay H, Topal T, Ozler M, et al. Persistence of hyperbaric oxygen-induced oxidative effects after exposure in rat brain cortex tissue. *Life Sci.* 2007; 80:2025-2029.
18. Oter S, Topal T, Sadir S, et al. Oxidative stress levels in rats following exposure to oxygen at 3 atm for 0-120 min. *Aviat Space Environ Med.* 2007; 78:1108-1113.
19. Kim CH, Choi H, Chun YS, Kim GT, Park JW, Kim MS. Hyperbaric oxygenation pretreatment induces catalase and reduces infarct size in ischemic rat myocardium. *Pflugers Arch.* 2001; 442:519-525.
20. Dennog C, Radermacher P, Barnett YA, Speit G. Antioxidant status in humans after exposure to hyperbaric oxygen. *Mutat Res.* 1999; 428:83-89.
21. Yogarathnam JZ, Laden G, Madden LA, et al. Hyperbaric oxygen: a new drug in myocardial revascularization and protection? *Cardiovasc Revasc Med.* 2006; 7:146-154. Review.
22. Rossignol DA, Rossignol LW, James SJ, Melnyk S, Mumper E. The effects of hyperbaric oxygen therapy on oxidative stress, inflammation, and symptoms in children with autism: an open-label pilot study. *BMC Pediatr.* 2007; 7:36.
23. Bader N, Bony-Westphal A, Koch A, Rimbach G, Weimann A, Poulsen HE, Müller MJ. Effect of hyperbaric oxygen and vitamin C and E supplementation on biomarkers of oxidative stress in healthy men. *Br J Nutr* 2007; 98:826-833.
24. Benedetti S, Lamorgese A, Piersantelli M, Pagliarani S, Benvenuti F, Canestrari F. Oxidative stress and antioxidant status in patients undergoing prolonged exposure to hyperbaric oxygen. *Clin Biochem.* 2004; 37:312-317.
25. Bild DE, Selby JV, Sincock P, Browner WS, Braveman P, Showstack JA. Lower-extremity amputation in people with diabetes. *Epidemiology and prevention.* *Diabetes Care.* 1989; 12:24-31.
26. Aydin A, Orhan H, Sayal A, Ozata M, Sahin G, Isimer A. Oxidative stress and nitric oxide related parameters in type II diabetes mellitus: effects of glycemic control. *Clin Biochem.* 2001; 34:65-70.
27. Cabigas BP, Su J, Hutchins W, et al. Hyperoxic and hyperbaric-induced cardioprotection: role of nitric oxide synthase 3. *Cardiovasc Res.* 2006; 72:143-151.
28. Hink J, Thom SR, Simonsen U, Rubin I, Jansen E. Vascular reactivity and endothelial NOS activity in rat thoracic aorta during and after hyperbaric oxygen exposure. *Am J Physiol Heart Circ Physiol* 2006; 291:1988-1998.
29. Chen SJ, Yu CT, Cheng YL, Yu SY, Lo HC. Effects of hyperbaric oxygen therapy on circulating interleukin-8, nitric oxide, and insulin-like growth factors in patients with type 2 diabetes mellitus. *Clin Biochem.* 2007; 40:30-36.

30. Sundaram RK, Bhaskar A, Vijayalingam S, Viswanathan M, Mohan R, Shanmugasundaram KR. Antioxidant status and lipid peroxidation in type II diabetes mellitus with and without complications. *Clin Sci (Lond)*. 1996; 90:255-260.
31. Srivatsan R, Das S, Gadde R, et al. Antioxidants and lipid peroxidation status in diabetic patients with and without complications. *Arch Iran Med*. 2009; 12:121-127.
32. Gürdöl F, Cimsit M, Oner-Iyidogan Y, Körpınar S, Yalçınkaya S, Koçak H. Early and late effects of hyperbaric oxygen treatment on oxidative stress parameters in diabetic patients. *Physiol Res*. 2008; 57:41-47.
33. Matsunami T, Sato Y, Sato T, Ariga S, Shimomura T, Yukawa M. Oxidative stress and gene expression of antioxidant enzymes in the streptozotocin-induced diabetic rats under hyperbaric oxygen exposure. *Int J Clin Exp Pathol*. 2010; 3: 177-188.
34. Yatsuzuka H. Effects of hyperbaric oxygen therapy on ischemic brain injury in dogs. *Masui*. 1991; 40:208-223.
35. Tepic S, Zivkovic M, Terzic N, Krivokuca R, Ljesevic B, Jakovljevic V. Effect of hyperbaric oxygen treatment on oxidative stress in patients having diabetes mellitus type 2. *Med Pregl*. 2009; 62:225-230.
36. Ekanayake L, Doolette D. Effects of hyperbaric oxygen treatment on blood sugar levels and insulin levels in diabetics. *SPUMS J*. 2001; 31:16-20.
37. Longoni C, Camporesi EM, Buizza M, et al. Reduction in insulin requirements during hyperbaric therapy. *Undersea Biomed Res*. 1998; 15:16-17.
38. Rose RE, Rice JH, Kraft KL, et al. An ongoing study of plasma glucose measurement in diabetic patients during hyperbaric oxygen therapy. *Undersea Hyperbar Med*. 2001; 28:32.
39. Nwafor TS, Collins N. Managing low blood glucose levels in patients undergoing hyperbaric oxygen therapy. *Ostomy Wound Manage*. 2014; 60:12-5.
40. Wilkinson D, Chapman IM, Heilbronn LK. Hyperbaric oxygen therapy improves peripheral insulin sensitivity in humans. *Diabet Med*. 2012; 29:986-969.
41. Al-Waili NS, Butler GJ, Beale J, et al. Influences of hyperbaric oxygen on blood pressure, heart rate and blood glucose levels in patients with diabetes mellitus and hypertension. *Arch Med Res*. 2006; 37:991-997.
42. Trytko B, Bennett MH. Blood sugar changes in diabetic patients undergoing hyperbaric oxygen therapy. *SPUMS J*. 2003; 33:62-69.
43. Karadurmus N, Sahin M, Tasci C, et al. Potential benefits of hyperbaric oxygen therapy on atherosclerosis and glycaemic control in patients with diabetic foot. *Pol J Endocrinol*. 2010; 61:275-279.
44. Berkovitch M, Tsadik R, Kozer E, Abu-Kishk I. The effect of hyperbaric oxygen therapy on kidneys in a rat model. *Scientific World Journal*. 2014; 2014:105069.
45. Rubinstein I, Abassi Z, Milman F et al. Hyperbaric oxygen treatment improves GFR in rats with ischaemia/reperfusion renal injury: a possible role for the antioxidant/oxidant balance in the ischaemic kidney. *Nephrology Dialysis Transplantation*. 2009; 24:428-436.

